

# CelCradle-500 Technical Report IV

## Cultivation of Sf-9 Insect cells and baculovirus expression system in SF 900 II media\*

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## 1 Description

CelCradle-500 provides a powerful tool to achieve high cell density and high productivity of target bioproducts in a cell culture because it has a unique feature of offering high oxygen transfer and low shear stress culture environment. Users can easily collect highly concentrated cells, virus or secreted products from one 500 ml CelCradle-500 bottle. In this study, the application of CelCradle-500 for growth of Sf-9 insect cells and production of baculovirus and enhanced GFP protein is illustrated. Three culture systems, i.e. spinner flask, spinner basket, and CelCradle, were compared simultaneously.  $1.5 \times 10^8$  Sf-9 insect cells were seeded in one CelCradle-500 unit. The cells were infected by BAE baculovirus with a MOI of 10. A total of  $3.45 \times 10^{12}$  baculovirus and 14,355 unit of e-GFP protein was produced in one CelCradle unit. e-GFP protein production in CelCradle system is 1.8 folds higher than a spinner basket and 7.9 folds higher than a 250 ml spinner flask. This technical sheet provides a general protocol for users to start up their culture. However, the optimum condition of each cell culture for each case may require the users to determine.

## 2 Material

Device	Cell Line	Medium	Seed	Virus	MOI
CelCradle-500, Spinner Flask (Corning), Spinner Basket (NBS)	Sf-9	SF900 II (Gibco)	$3.0 \times 10^5$ cells/bottle	BacCE baculovirus*	10

\*BacCE baculoviruses are kindly provided by Dr. Hu in Tsing-Hua University, Taiwan

## 3 Protocol for CelCradle System

### 3.1 Inoculum Preparation

Prepare one 250ml spinner flask and inoculate  $2.5 \times 10^5$  suspended cells/ml in 120ml SF 900 II culture media. Culture at 90 rpm, 28°C for 4 days. After cell density reaches above  $1.5 \times 10^6$  cells/ml and viability remain above 95%, it is ready for the preparation of inoculation. Collect  $1.5 \times 10^8$  suspended cells from the spinner flask by centrifugation and collect in one 50ml centrifuge tube with 50ml fresh media.

### 3.2 Preparation before cell seeding

Place CelCradle controller in a 28°C incubator. Set up the inoculation parameters (See below). Warm up SF900 II medium in 28°C water bath. Take out one CelCradle bottle aseptically and place it in a biosafety cabinet. Open the cap and add 450 ml fresh culture medium in the bottle.

### 3.3 Inoculation

Open the cap and dispense 50 ml media containing  $1.5 \times 10^8$  suspended cells that has been prepared previously on top of the matrices of CelCradle-500. Bring the bottle and lock up on the CelCradle controller in incubator at 28°C.

### 3.4 Culture

Press “START” button to start the controller. After 2 to 3 hours, reset the parameters for culture condition. Usually, above 90% cells will be immobilized in the matrices within 30 minutes. The inoculation parameters are set as below:

Rising rate	Top Holding Time	Down Rate	Bottom Holding Time
2.0 mm/s	20 sec	2.0 mm/s	0 sec

The culture parameters are set as below:

Rising rate	Top Holding Time	Down Rate	Bottom Holding Time
2.0 mm/s	3.5 10 sec	2.0 mm/s	1 min 30 sec

Monitor the pH, residual glucose concentration and other metabolic in order to predict the time for virus infection or the termination of culture.

When pH below 6.0 during culture, add Bis-Tris to re-adjust the pH to above 6.5. There is no need to replenish media at this application.

*The setup parameters are only for reference. It does not necessary to be optimum parameters.*

### 3.5 Infection

When the growth reach mid-log phase, where GUR is 25 mg/hr, prepare for virus infection. Replace the culture medium and Infect cells with MOI=10. Control the pH by adding Bis-Tris to above 6.5.

## 4 Result

Below shows the result of cell growth, cell density, virus production, and E-GFP protein production in the CelCradle-500 system. Figure 1 shows the profile of glucose uptake rate in three different culture systems. CelCradle-500 provides optimum environment for cells to grow up to  $1.4 \times 10^7$  cells/ml, while other two systems have relatively lower cell density.

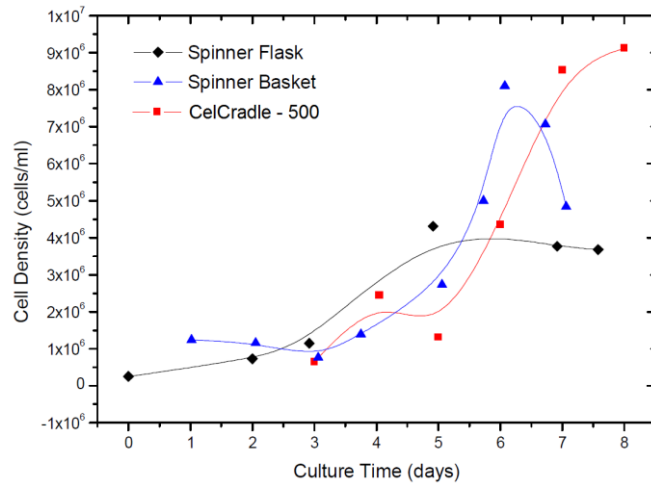


Figure 1. Profile of cell density in three different culture systems

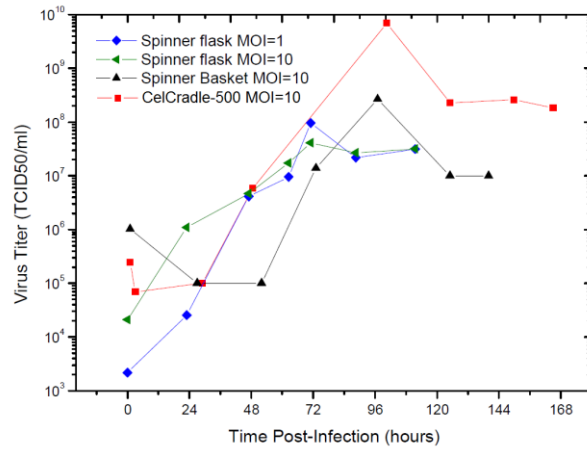


Figure 2. The baculovirus production in three different culture system. Virus productivity in CelCradle-500 is 30 folds higher than spinner basket system and 240 folds higher than spinner flask system.

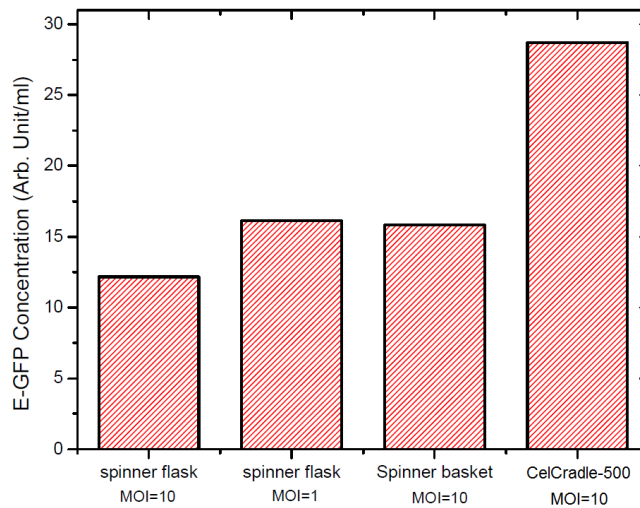


Figure 3: The E-GFP productivity in three different system. Protein concentration in CelCradle-500 is about 2 folds higher compared with other systems.

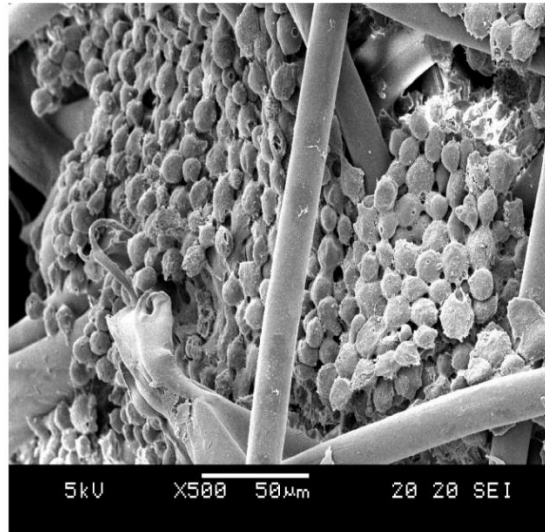


Figure 4: The SEM photo of S-9 cells cultivated in BioNOC II matrix in CelCradle-500 system

The result indicates that CelCradle cell culture system can be applied in Sf-9 insect cell culture for high cell density culture and high viruses and protein production.

## 5 Summary

Below is the summary of the comparison of sf-9 cell growth and baculovirus/protein production in various lab culture systems.

System Used	Spinner flask	Spinner Basket / Fibracel disk	CelCradle / BioNOC matrices
Description	250 ml flask/150 ml medium	1000cm <sup>3</sup> bottle / 450ml medium / 100cm <sup>3</sup> (10g) carriers	1000 ml bottle / 500 ml medium / 100cm <sup>3</sup> (10g) carriers
Total surface area (cm <sup>2</sup> )	None	12,000	13,000
Mode of Operation	None	Batch	Batch

<b>Medium for growth/infection</b>	SF 900 II	SF 900 II	SF 900 II
<b>Max. cell density (cells/ml of medium)</b>	$4.3 \times 10^6$	$8.1 \times 10^6$	$1.4 \times 10^7$
<b>Total Cell Number</b>	$6.5 \times 10^8$	$3.6 \times 10^9$	$7.2 \times 10^9$
<b>Multiplicity of infection (MOI)</b>	1 and 10	10	10
<b>Virus productivity (TCID<sub>50</sub>/ml)</b>	$9.6 \times 10^7$	$2.6 \times 10^8$	$6.9 \times 10^9$
<b>Total virus production (TCID<sub>50</sub>)</b>	$1.44 \times 10^{10}$	$1.17 \times 10^{11}$	$3.45 \times 10^{12}$
<b>E-GFP (arb. U/ml)</b>	16.13 (MOI=1) and 12.16 (MOI=10)	17.62	28.71
<b>Total E-GFP (arb. U)</b>	2419.6 (MOI=1) and 1824.03 (MOI=10)	7927	14355

## 6 VacciXcell Technical Support

For queries and comments, please contact the VacciXcell Technical Support team.

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